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Test Request / Chain of Custody

Reporting and Billing Information

Client: Sample:

Client / Operation:	
Contact:	
Report Address:	
Billing Address:	
Tel	Fax
Quote/PO/Job	

Rush: 50% surcharge; 100% surcharge (evenings and weekends); Microtox® required in less than 12 hours 100% surcharge; 24 hours 50%

Tests Requested (codes on back)
 (example: trout with 5 treatments, TR-D)

Sample Received Intact (Y / n)

Notes: S = single treatment, D = multiple treatments

Check appropriate box below

Sample ID	Sampled By / Date / Time	Location	Method	Type

Relinquished By	Date / Time
<input style="width: 95%;" type="text"/>	<input style="width: 95%;" type="text"/>

Received By (HQ)	Date / Time
<input style="width: 95%;" type="text"/>	<input style="width: 95%;" type="text"/>

Our liability is limited to the cost of the test requested on the sample as received. No liability in whole or in part is assumed for the collection, handling or transport of the sample, application or interpretation of the test data or results in part or in whole.



Test Codes, Volumes, and Sampling Guide

Test (call lab for complete list)	Code	Sample Volume (L)		Holding Time (days)
		undiluted (S)	multiple (D)	
Freshwater <i>Daphnia magna</i> (48 hours, static acute) rainbow trout (96 hours, static acute) algal growth inhibition (72 hours, static) <i>Ceriodaphnia</i> survival and reproduction (7 day) * fathead minnow survival and growth (7 day) * <i>Lemna minor</i> survival and growth (7 day)* Salmonid early life stage	DA	1	1	5
	TR	20	40	5
	AG	1	1	3
	CD	3	6	3
	FM	10	20	3
	LM	1	1	3
	ELS	80	160	3
	HA	2	5	42
	CT	2	5	42
	LV	2	5	42
Sediments <i>Hyalella azteca</i> (14 day, static) chironomids (10 day, static) <i>Lumbriculus</i> (10 day, static) marine amphipod or polychaete heterotrophic/hydrocarbon degrading bacteria fungal enumeration	AM/PO	2	5	42
	THB/HDB	0.25	0.25	na
	FE	0.25	0.25	na
	ME	0.25	0.25	na
	TC/EC,FC	0.25	0.25	1
	BOD	1	1	1
	CGE		call lab	96
	PPI	0.1		na
	AGID		0.1	na
	G50/BL	0.1	0.1	3
Other seedling emergence root elongation earthworm survival (14 days) treatability ecological health indices total suspended solids earthworm chronic (60 days) plant growth studies G50 Microtox / bacterial luminescence	SE	1	1	na
	RE	1	1	na
	EW	1	1	na
	TREAT		call lab	na
	EHI		call lab	na
	TSS		call lab	1
	EW		call lab	na
	PGT		call lab	na

Notes: S, single treatment; D, definitive; na, not applicable; * sample must be divided equally into 3 separate containers upon collection or at lab
For marine testing call Kate de Windt or Ingrid Carleton-Dodds

Sampling and Shipping Instructions (Supporting Work Instruction 4.3.1.4.1)

Containers

liquids
glass, polyethylene, or polypropylene container
amber glass bottle with teflon lined cap is preferred for small samples (<2L)
collapsible 6 and 20L carboys or plastic pails and gasket lined lids
glass, polyethylene, or polypropylene container (wide mouth)
polyethylene bags (6mL)
air
new Ziploc® bags for viable samples (RCS), spore traps, swab and tape samples on slides
new ziploc® bags or jars for bulk samples

Sampling

liquids
rinse three times with sample before filling (>5% of container volume for each rinse)
fill container to exclude air (minimal headspace) and seal with cap; cover cap with tape and initial
keep composited sample cool during collection (4 ± 1 °C)
solids
fill and seal container
air
follow manufacturers instructions for air sampling devices
take bulk samples with sterile cutting tool
take tape samples with 5 cm piece of clear tape, place on surface to be sampled, remove and affix to clean slide (avoid creases)

Labelling

all
label sample container with type of sample (effluent, groundwater, surface water, etc.), source, date, time of collection, and name of sampler (s)
tape lids of containers and initial tape - fill out chain of custody and ship with sample

Transport

samples should be transported in the dark, at 1-8°C. During the winter season, ship samples in a heated truck, in order to keep them from freezing.